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## FREQUENCY OF RICKETTSIA SPS. IN

## DERMACENTOR VARIABILIS AND AMBLYOMMA AMERICANUM IN CENTRAL HANOVER COUNTY, VIRGINIA

Ву

PEGGY ANN KEEFE

B.S., UNIVERSITY OF UTAH, 1982

A Thesis

Submitted to the Graduate Faculty

of the University of Richmond

in Candidacy

for the degree of

MASTER OF SCIENCE

in

Biology

August, 1989

Richmond, Virginia

UNIVERSITY OF RICHMOND VIRGINIA 23173

## FREQUENCY OF <u>RICKETTSIA</u> SPS. IN <u>DERMACENTOR VARIABILIS</u> AND <u>AMBLYOMMA</u> <u>AMERICANUM</u> IN CENTRAL HANOVER COUNTY, VIRGINIA

Ву

PEGGY ANN KEEFE

APPROVED:

W. COMMITTEE CHAT

COMMITTEE MEMBERS

EXAMINING COMMITTEE:

## Acknowledgements

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## TABLE OF CONTENTS

ACKNOWLEDGEMENTSii
INTRODUCTIONl
MATERIALS AND METHODSl
SITE DESCRIPTIONS2
RESULTS
DISCUSSION
CONCLUSION
FIGURE9
TABLES10
LITERATURE CITED12
APPENDICES15
VITA

#### INTRODUCTION

In the eastern United States, <u>Dermacentor variabilis</u> is the primary vector of <u>Rickettsia</u> <u>rickettsii</u>, the etiological agent for Rocky Mountain Spotted Fever. Cases of Rocky Mountain Spotted Fever in eastern United States increased steadily from 1959 to 1979 to a peak of one case per 200,000 people in 1979 (<u>Morbidity and Mortalty Weekly Report</u>, 1985). In 1976, 98.9% of the reported cases of Rocky Mountain Spotted Fever were east of the 100th meridian with 1.1% occurring west of the 100th meridian (Burgdorfer, 1977). Hanover County, Virginia is an endemic area for this disease (Sonenshine, 1971); the last reported case was in 1984 (Personal communication, Dept. of Epidemiology, Va., 1989). The present study was to determine if rickettsial infection was present in the tick population at three sites in Hanover County, Virginia.

## MATERIALS AND METHODS

Three study sites were chosen in Hanover County, Virginia, an area endemic for Rocky Mountain Spotted Fever. In 1988, from April to September, in a total of 38 collection days, sites were flagged with a 142 cm x 142 cm white flannel cloth attached to a yardstick (Benach et al., 1977). The cloth was dragged over the vegetation and ticks that adhered were removed with tweezers and placed in a small vial. Ticks were kept alive on moist cotton until a hemolymph test was performed. Sites usually were flagged in the late morning or early afternoon to avoid morning dew. Ticks also were obtained by removing them from humans and dogs. Ticks were identified following Sonenshine (1979).

The establishment of rickettsial infection was determined by the hemolymph test (Burgdorfer, 1970). The distal portion of the third leg of the tick was amputated with a scalpel and a drop of hemolymph placed on a slide. The slide was air dried, fixed with heat, stained with the Gimenez method and examined under oil immersion (1,600 x). Rickettsiae are primarily located within the cytoplasm of hemocytes, which stains green whereas the rickettsiae appear as pink or red spheres (Burgdorfer, 1970). The species of rickettsiae cannot be determined with this method.

#### SITE DESCRIPTIONS

Site 1 was a 119 m stretch of access road to a sewer pump station off of Atlee Station Road, 5 km southeast of Ashland, Virginia and 3.06 km from the intersection of Route 301 and Atlee Station Road. (Figure 1). A 1 m wide area of brush and grass on both sides of the road was flagged (Benach et al., 1977). Site 1 A was an additional 99 m of road added to the area in site 1. Ground vegetation consisted of grasses (Family: Gramineae) growing to an approximate height of 1 m. Trees, mainly white oak (Quercus alba) and pine (Pinus sp.) with a scattering of

beech (Fagus grandifolia), hickory (Carya sp.) and holly (Ilex opaca), provided a canopy over the road. During April and May, large sections of the low lying ground adjacent to the road were covered with water.

Site 2 was a 320 m stretch of a private gravel road in a five year old housing development off of Route 301, 11 km southeast of Ashland, Virginia and 0.8 km west of the end of Route 752 (Figure 1). A 0.5 m wide area on the north side of the road was flagged. Ground vegetation consisted of grasses and huckleberry (<u>Gaylussacia</u> sp.) bushes, reaching a height of 0.3 to 0.5 m. Trees along the north side of the road were predominantly white oak and pine with some holly and dogwood (Cornus florida).

Site 3 was a 63 m stretch along an abandoned, overgrown road on a private farm off of Atlee Station Road, 4.5 km southeast of Ashland, Virginia and 4.7 km from the intersection of Route 301 and Atlee Station Road (Figure 1). A 0.5 m wide area on both sides of the road was flagged. Vegetation consisted of honeysuckle vines (Lonicera sp.) and periwinkle (Vinca sp.), approximately 0.3 m tall with white oak and pine trees overhanging the road.

### RESULTS

A total of 388 ticks was collected by flagging at the three sites from April to September (Table 1). The predominant

species of tick at all three sites was <u>Dermacentor</u> <u>variabilis</u> (94%, 85%, and 75%). Site 3 had the highest incidence of <u>Amblyomma americanum</u> (25%) and site 2 was the only location where an Ixodes sp. tick occurred.

Hemolymph tests for the presence of rickettsiae (Burgdorfer, 1970) showed that site 2 was the only area where the organism was found. The incidence rate was 2.68%, where three female <u>D</u>. <u>variabilis</u> and one female <u>A</u>. <u>americanum</u> were infected (Table 2). The species of rickettsiae was not determined.

Of the 388 ticks collected, 16 had bacillary organisms present intracellulary and extracellularly in their tissues. The bacteria stained red with the hemolymph test. Two of the 16 ticks were collected at site 2; 14 were at site 1. <u>Dermacentor variabilis</u> composed 7.7% of the infected ticks and A. americanum 1.3% (Table 3).

### DISCUSSION

The distribution of ticks in this study is consistent with studies by Sonenshine (1971) in Virginia. Of the 3323 ticks collected from animal hosts in his work 97.8% were <u>D. variabilis</u> and 2.2% were <u>A. americanum</u> which is similar to the results in the present study except at site 3 where 75% were <u>D. variabilis</u> and 25% <u>A. americanum</u>. The high percentage of <u>A. americanum</u> at site 3 is not significant (P=0.5) and may be the result of a small data pool of only 15 ticks, five of which were <u>A</u>. <u>americanum</u> and ten were <u>D</u>. <u>variabilis</u> (See Appendix I). <u>Amblyomma americanum</u> is known to secrete aggregation pheromones (Harwood and James, 1979), and it is possible this influenced the clustering of these ticks in the same spot on the same day.

The 2.68% incidence of rickettsiae in this study is similar to results reported in other endemic areas. For example, Burgdorfer (1974) found a 3.5% incidence of rickettsiae in <u>D. variabilis</u> in a study in the Tennessee Valley Region; in South Carolina, Burgdorfer et al. (1975) reported a 4% incidence of rickettsiae in <u>D. variabilis</u> and Magnarelli (1979) found a 2.9% incidence of rickettsiae in <u>D. variabilis</u> in Connecticut.

Small mammals, e. g. meadow mice and rabbits, are primary hosts for the larval and nymphal stages of <u>D</u>. <u>variabilis</u> and <u>A</u>. <u>americanum</u> (McDade and Newhouse, 1986). After molting, the adult tick attaches to larger animals such as dogs, raccoons, foxes, and deer (McDade and Newhouse, 1986). Since rickettsiae, were found only at site 2, a possible explanation might relate to habitat disturbance. In 1980, a housing development was started at this site. In undisturbed areas, like site 1 and site 3, fewer small mammals should be present and a lesser chance for immature ticks to feed on rickettsemic hosts would be expected. In a disturbed area, i.e. site 2, more small mammals were expected because of an increase in suitable habitat (Pianka, 1979), thus

raising the chance of an immature tick feeding on a rickettsemic host and acquiting the rickettsiae.

All four ticks harboring the rickettsiae were females. The apparent sex bias in harboring rickettsiae reflects a small data pool as both male and female D. variabilis are equally susceptible to the rickettsial infection (Ricketts, 1907). Rickettsia rickettsii has never been recovered from a naturally infected A. americanum but rickettsiae related to the spotted fever group have been identified in this species (Burgdorfer et al., 1974). These rickettsiae react with anti-R. rickettsii conjugates in a fluorescent stain but do not produce a detectable infection (high fever or scrotal swellings) in laboratory animals (Burgdorfer et al., 1974). These rickettsiae may not be pathogenic to man, as a one year old boy in South Carolina, who was bitten by an A. americanum infected with this species, did not become ill (Burgdorfer et al., 1975).

At site 1, bacillary organisms were present at 7.7%, a frequency comparable to the 8.9% observed by Burgdorfer et al. (1974) in the Tennessee Valley Region. In South Carolina, bacillary organisms were discovered in 25 <u>D</u>. <u>variabilis</u> (Burgdorfer and Brinton, 1975). These organisms produced an infection throughout the tick and were passed transovarianly by females to their progeny. They were not pathogenic to laboratory animals nor to humans (Burgdorfer et al., 1975).

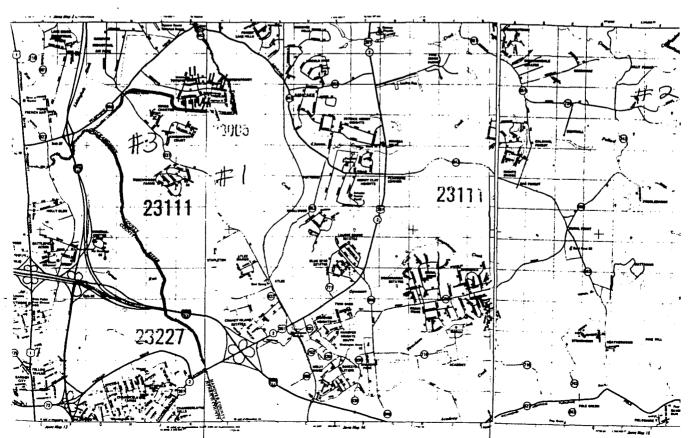
These bacillary organisms might be an inhibiting factor for rickettsial distribution at site 1. It has been shown that once a tick is infected with a species of rickettsia, it will not retain or accept a second species (Burgdorfer et al., 1981). Burgdorfer et al. (1981) discovered that the "East side agent", a rickettsia found only on the eastern side of the Bitter Root Valley, an area free of Rocky Mountain Spotted Fever, was retained by <u>D</u>. <u>variabilis</u> when challenged with <u>R</u>. <u>rickettsii</u>. The competition between <u>R</u>. <u>rickettsii</u> and the "East side agent" for tick hosts is thought to be responsible for relegating <u>R</u>. <u>rickettsii</u> to the western side of the Bitter Root Valley, Montana.

At site 1, where no rickettsiae were found, 7.7% of all ticks collected harbored the bacillary organism. At site 2, where there was a 2.68% incidence of rickettsiae, only 1.3% of the ticks collected had the bacillary organism. Possibly the bacillary organism was competing with the rickettsiae for tick hosts at site 1, limiting the distribution of rickettsiae. Only further study similar to that conducted with the "East side agent" can show if the bacillary organism is a factor in rickettsial distribution at these sites.

### CONCLUSION

The incidence of Rocky Mountain Spotted Fever in Hanover County is similar to other endemic areas. Of the 388 ticks collected, four ticks has a rickettsia present in their tissues. There was a 2.68% infection rate at site 2 which could be the result of an increased small mammal population, a result of a disturbed habitat. The bacillary organism found in 7.7% of <u>D</u>. <u>variabilis</u> collected at site 1 might be competing with rickettsiae for hosts and may be a limiting factor in the distribution of rickettsiae to certain areas of Hanover County, Virginia. Figure 1. a) Map of Hanover County, Virginia; The three test sites are located within the red square. b) Enlargement for area in red square (a). Three test sites are designated by red numbers.





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Site	Dermac variab			omma Lcanum	Ixod	
	NO.	90 	No.	98 98	No.	8
1	171	94	11	6	0	0
l A	36	97	1	3	0	0
2	126	84.5	22	14.8	1	0.67
3	15	75	5	25	0	0

Table l.	Distribution of tick species collected by	flagging
	at test sites in Hanover County, Virginia	from
	April to September 1988.	

Table 2. Frequency of ticks infected with <u>Rickettsia</u> spp. collected by flagging in Hanover <u>County</u>, Virginia from April to September of 1988.

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Site	Rickett No.	sia spp. %
1	0	0
1 A .	0	0
2	4	2.68
3	 0	0

			· .	
Site	Bacill No.	i Present	Dermacentor variabilis No.	Amblyomma americanu No.
1	14	7.7	. 11.	3
1 A	0	0	0	0
2	2	1.3	2	0
3	0	0	0	کل ا

Table	3.	Frequency	of	ticks :	infected	with	bacil	llary	organisms,
		collected	in	Hanover	r County	, Virg	ginia	from	April
				to Ser	ptember ]	1988.			

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- Appendix I. Collection data from test sites in Hanover County, Virginia from April to September, 1988.
  - + present
  - - absent
  - \* Bacillary organism present
  - ₭ Hemolymph test positive

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υττε Stage	A	Ą	A	A	A	A	A	A	A	A	A	A	A	A	A
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Flagging	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
apecimen # asmissed2	06	91	92	93	. 94	95	96	97	86	66	100	101	102	103	104
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#1	specimen #	105	106	107	108	109	110	111	1,12	113	114	115	116	117	. 118	119	
n Site: .	ЭдбŪ	5/31/88	6/11/88			·	<i>`</i>	•		6/12/88				••••••			
Location Site:	anoitibnoD		10,00 a.m.	Cool						24	82 F, Sunny, Clear		· · ·	. //			

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#7	# nemiceq	120	121	122	123	124	125	126	127	128	129	130	131	132	133	134	
ı Site:	Dаte	6/12/88		6/13/88					6/15/88				6/17/88	6/23/88			
Location Site:	snoitibnoD	· · · · · · · · · · · · · · · · · · ·		8:20 am, 64 <sup>0</sup> F Shadv, Cool	1				11:00 am, 84 <sup>0</sup> F Sunny	·········				9:00 am, 94 <sup>0</sup> F Hot			

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Location Site:	Ðàtg	6/23/88						6/27/88					6/30/88		7/2/88	7/4/88
catio		,				-	-	, 72 <sup>0</sup> F		_	•		680F			
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#1	# asmissq2	150	151	152	153	154	155	156	157	158	159	160	161	162	163	164
Location Site:	Date	7/15/88		7/16/88						• •	7/17/88					a.m.,88 <sup>9</sup> 7/18/88
Locatio	гиоітібиоЭ	11:30 a.m.,	94 F, HOT	10;15 a.m.,							11:00 a.m.,	ot r, Warm	• · ·			10:15 a.m.,88

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	Dog									+	+	+	+			
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1	Flagging	+	+	+	+		+	+	+					+	+	+
#1	# nemiceq2	1.65	166	167	168	. 169	170	171	172	173	174	175	176	177	178	179
n Site:	Date	7/18/88			7/21/88	7/24/88	7/25/88	7/27/88						880F 7/29/88	8/3/88	90. <sup>0</sup> F 8/8/88
Location Site:	гиоі ті био)				9:00 am, 75 <sup>0</sup> F	TOOD ' ADUTM	1:30 pm, 850F Wet Warm	12:30 pm, 820F Wet						am, bad	11:30 am, 82°F 8 Cloudv	ц Ц

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wsŗ	Bacillary Organ			1										
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ļ	Flagging	+	.+	+	+	+	+	+	+	+	+	+	+	+	+	+
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#1 A	# nemiced	31	32	33	34	. 35	36	37						
n Site: _	Date	<b></b> •	1				8/13/88	 8/17/88						
Location Site:	anoitibnoJ	Cool, Shady					6:45 p.m.,	10:30 a.m.,	Hot					

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#2	# nemiceqS	1	2	£	4	2	9	7	8	6	10	11	12	13	14	15
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	Dog															
	nsmuH													8.		
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#2	# nemiceq2	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60
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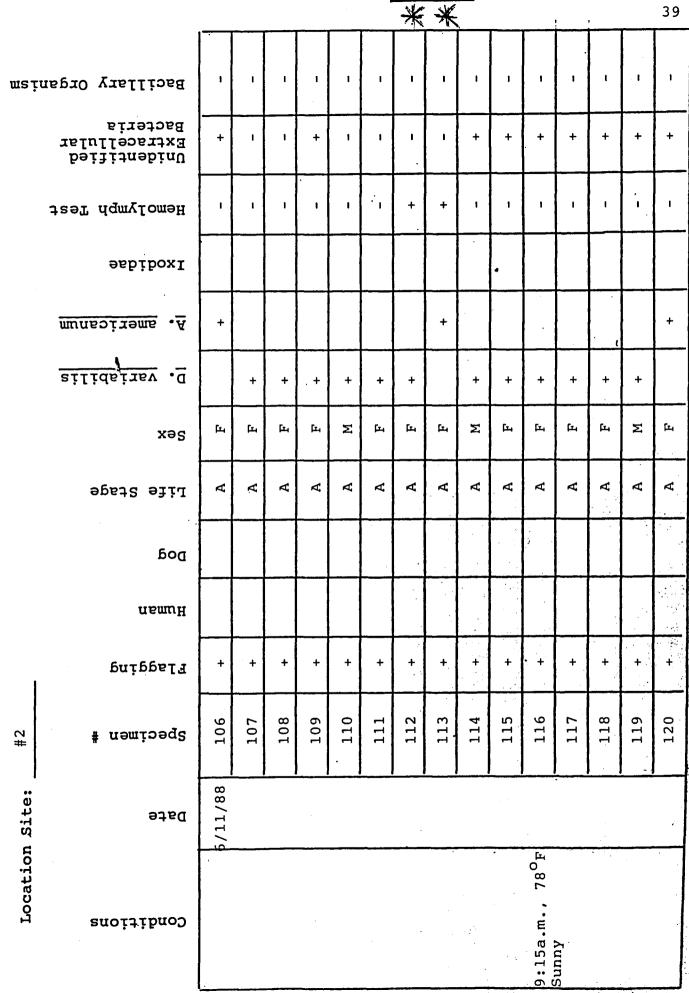
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Bacillary Organism t 1 1 1 I. L t ŧ 1 1 1 I. I ŧ t Вастетіа Unidentified Extracellular + ÷ ι + + + L 1 L + t + I. L ÷ i ÷1 t ł L I. 1 I. Ł L I. I t I I. Hemolymph Test ssbiboxl + A. americanum + + D. Variabilis + + + + + + + + + +. <del>+</del> + xəs ኴ Гц Ē ш ſ۳ı Гч Бч Бц Σ Ŀι Σ Бч Гч Гч A A A А A 4 4 A z 4 A A 4 4 A Life Stage Dog nemuH Flagging + ÷ + + + + + + + ÷ + + ÷ + + 66 68 70 .73 75 62 64 71 72 63 65 67 69 74 61 # uemiced 5/31/88 5/28/88 Date , p.m., 200111bnoD ы 90 90 -1 - 7

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1	Flagging	+	+	+	+	+	+	+	+	+	+	• +	+	+	+	+
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Presented here is the original research proposal for this work. Unfortunately due to restrictions found in the federal guidelines, these experiments could not be attempted. The fluorescent antibody stain for spotted fever group rickettsiae could not be obtained from Centers for Disease Control in Atlanta. A similar problem occurred in getting <u>Peromyscus</u> antibodies for the IFA test for <u>R</u>. <u>rickettsii</u> and <u>R</u>. <u>montana</u> antibodies in the field mice captured in this study. So instead of an immunity demonstration, the present study focused on a survey of rickettsial infection at the Three test sites in Hanover County, Virginia.

The goal of this research project is to ascertain whether an infection of <u>Rickettsia montana</u> in <u>Microtus pennsylvanicus</u>, the meadow vole and <u>Peromyscus polionotus</u>, the white footed mouse will impart an immunity to an infection of <u>Rickettsia</u> <u>rickettsii</u>, the etiological agent for Rocky Mountain Spotted Fever. <u>Rickettsia montana</u> is highly pathogenic to meadow voles and could increase the likelihood of immunity against R. rickettsii.

The incidence of Rocky Mountain Spotted Fever in humans has decreased in the West over the past few decades to a very low frequency (Burgdorfer, 1977). Infection in the tick is passed transovarianly for only eight generations and new rickettsia must be obtained from rickettsemic hosts like the meadow vole or the white footed mouse. If these hosts are becoming immune to <u>R</u>. <u>rickettsii</u>, the ticks will not be reinfected and the pathway of transmission will be broken.

Three test sites have been selected in Hanover County, Virginia, an area known for Rocky Mountain Spotted Fever. The last reported case was in 1984 (Dept. of Epidemiology, Virginia).

The experimental design is in three parts. First is to collect ticks in these areas to determine the frequency of infection by <u>R</u>. <u>montana</u> and <u>R</u>. <u>rickettsii</u>. Collection will be by flagging and removal from host animals. The hemolymph test will be used to show the presence of rickettsiae and a fluorescien isothiocyanate labelled antibody stain will be used to identify the rickettsiae as part of the Spotted Fever Group. This method is unable to identify species.

The second part of the experimental design is to collect meadow voles and white footed mice, obtain blood samples and determine the frequency of infection by these two rickettsiae. Specimens will be collected in Sherman life traps, anesthesized with ether, and blood drawn from the orbital sinus. After marking for identification, animals will be released to their natural environment. Blood samples will undergo the indirect

fluorescent antibody or IFA test that will identify the presence of antibodies to <u>R</u>. montana and <u>R</u>. rickettsii. This is a species specific test.

The last part of the experiment is to conduct an immunity challenge test. Laboratory mice will be infected with the M/5-6 strain of <u>R</u>. <u>montana</u>. After two weeks or the establishment of an antibody titer, the mice will be infected with the "Sawtooth-+2" strain of <u>R</u>. <u>rickettsii</u>. Injections will be intraperitoneally and .1 cm<sup>3</sup> of an infectious yolk sac suspension diluted with brain hear infusion broth will be used. Temperatures will be taken daily and other signs of clinical infection observed. Titers will be measured with the IFA test. If the infection of <u>R</u>. <u>montana</u> has given an immunity against infection of <u>R</u>. <u>rickettsii</u>, the mice should not have severe clinical symptoms. If immunity is not present, the mice will probably die from the infection of <u>R</u>. <u>rickettsii</u> since this is a very virulent strain. Peggy Ann Keefe was born in Chillicothe, Ohio, April 7, 1959. She attended grammar school in Orlando, Florida and high school in Salt Lake City, Utah. A Bachelor of Science degree in biology was received from the University of Utah in 1982. From 1983 to 1986, Miss Keefe was employed at Native Plants, Inc. where she was responsible for producing chemically induced mutations in commerical strawberries and grapes. She completed requirements for a Master of Science degree in biology at the University of Richmond in August, 1989. While at the University of Richmond, she was a member of Beta Beta Beta and an associate member of Sigma Xi. Miss Keefe is currently engaged in small cell lung cancer research at the Veterans Administration Medical Center, Richmond, Virginia.

Vita